LETTER TO THE EDITOR

DETECTION OF THE MCR-1 GENE IN BACTERIAEMIA CAUSED BY Escherichia coli AND Klebsiella pneumoniae

Coralith García^{1,2,a}, Lizeth Astocondor^{1,b}, Noemi Hinostroza^{1,c}, Fiorella Krapp^{1,d}, Jan Jacobs^{3,e}

- ¹ Alexander von Humboldt Institute of Tropical Medicine, Universidad Peruana Cayetano Heredia. Lima, Peru.
- ² Cayetano Heredia National Hospital. Lima, Peru.
- ³ Antwerp Institute of Tropical Medicine. Antwerp, Belgium
- ^a Physician, PhD in Biomedical and Pharmaceutical Sciences, ^b licentiate in Medical Technology, ^c licentiate in Biology, d physician specialized in Infectious Diseases, Master in Sciences, ^c Microbiologist.

To the Editor. Since its emergence, isolates of carbapenem-resistant *Klebsiella pneumoniae* have been disseminated in hospitals in different regions of Peru. Currently, carbapenemase type New Delhi metallo-beta-lactamase (NDM) is the most frequently detected among *K. pneumoniae* and *Escherichia coli* isolates ⁽¹⁾. In these infections, colistin is often the antibiotic of last resort, since new antibiotics against these pathogens are not included in Peru's National Drug Formulary.

In order to assess colistin susceptibility, the recommended method for estimating the minimum inhibitory concentration is broth microdilution, but this method is not practical for most microbiology laboratories. In contrast, the disk elution method uses reagents and equipment usually available in routine laboratory processes and is less time consuming. This method is also recommended by the Clinical and Laboratory Standards Institute (CLSI) for the evaluation of colistin in *Enterobacteriaceae*⁽²⁾.

Correspondence. Coralith García Apac, coralith.garcia@upch.pe

Received. 11/30/2023 Approved. 04/17/2024 Online. 27/05/2024



This work is licensed under a Creative Commons Attribution 4.0 International

Copyright © 2024, Revista Peruana de Medicina Experimental y Salud Pública The mechanisms of colistin resistance among gram-negative bacilli include mainly resistance genes located in chromosomes or in transferable elements (plasmids or transposons), the latter being the most widespread mechanism. The *mcr*-1 gene, including its variants and sub-variants that are carried on these plasmids or transposons have been detected worldwide among several species of gram-negative bacteria either in humans, animals or in the environment. The *mcr* genes encode a transferase that increases the cationic charge of the lipopolysaccharide causing decreased binding of colistin to the lipopolysaccharide ⁽³⁾.

We evaluated colistin susceptibility in 317 isolates of E. coli (n=199) and K. pneumoniae (n=118) obtained from blood cultures. These samples were collected during an antimicrobial resistance surveillance study that included 15 hospitals in 12 regions of Peru between 2017-2019 (4). Bacterial identification was performed through conventional biochemical tests and susceptibility to carbapenems tests were performed through the disc-diffusion method. In order to assess susceptibility to colistin, the disk elution technique was performed following the procedures described by CLSI (2), identifying 9.1% (29/317) isolates resistant to colistin ($\geq 4 \mu g/mL$), of which 7.5% (15/199) and 11.9% (14/118) were E. coli and K. pneumoniae isolates, respectively (Table 1). Regarding resistance to carbapenems, 11.0% of K. pneumoniae isolates were resistant to some carbapenemics, on the other hand, none of the E. coli isolates were found to be resistant

We evaluated the presence of the *mcr*-1 gene by conventional PCR technique in all colistin-resistant isolates. The primers CLR-5F and CLR-5R described by Liu *et al.* ⁽⁵⁾ were used. The master mix was prepared to a final volume of 25 µl with 0.2 µM of each primer, 0.2 mM dNTP, 1.5 mM MgCl2, 1X buffer, 0.03 U/µl of GOTAQ[®] G2 FLEXI DNA polymerase (Promega, Madison, USA) and 2 µl of DNA. The cycling conditions described by Faccone *et al.* were used with a modification in the hybridization temperature of 72 °C for 5 min. The size of the amplicons was 309 base pairs.

The *mcr*-1 gene was detected in 9 of the 29 colistin-resistant isolates, 8/15 in *E. coli* isolates and 1/14 in *K. pneumoniae* isolates (Table 1). These nine isolates were susceptible to carbapenems and were distributed in 7 of the 15 hospitals in 5 of the 12 evaluated regions (Lima, La Libertad, Ancash, Loreto and Puerto Maldonado).

The epidemiology of *Enterobacteriaceae* species carrying the *mcr*-1 gene in Peru and other Latin American countries may be underestimated since colistin resistance testing is not routinely performed on all gram-negative bacteria, and in

Cite as. Garcia C, Astocondor L, Hinostroza N, Krapp F, Jacobs J. Detection of the mcr-1 gene in bacteriemia caused by *Escherichia coli* and *Klebsiella pneumoniae*. Rev Peru Med Exp Public Health. 2024;41(2):223-4. doi: 10.17843/rpmesp.2024.412.13507.

	Region	Escherichia coli					Klebsiella pneumoniae				
Hospital		N	Colistin resistance		Detection of <i>mcr</i> -1 gene		N	Colistin resistance		Detection of <i>mcr</i> -1 gene	
			n	(%)	n	(%)		n	(%)	n	(%)
1	Lima	88	6	(6.8)	1	(1.1)	47	6	(12.8)	0	(0.0)
2	Lima	23	3	(13.0)	3	(13.0)	9	1	(11.1)	0	(0.0)
3	Lima	17	1	(5.9)	0	(0.0)	19	2	(10.5)	0	(0.0)
4	La Libertad	16	1	(6.3)	1	(6.3)	6	1	(16.7)	0	(0.0)
5	La Libertad	12	1	(8.3)	1	(8.3)	8	0	(0.0)	-	-
6	Cusco	9	0	(0.0)	-	-	4	0	(0.0)	-	-
7	Arequipa	8	1	(12.5)	0	(0.0)	5	0	(0.0)	-	-
8	Madre de Dios	6	1	(16.7)	1	(16.7)	2	0	(0.0)	-	-
9	Tacna	5	0	(0.0)	-	-	2	1	(50.0)	0	(0.0)
10	Loreto	4	0	(0.0)	-	-	6	3	(50.0)	1	(16.7)
11	Ancash	4	1	(25.0)	1	(25.0)	2	0	(0.0)	-	-
12	Ucayali	3	0	(0.0)	-	-	0	0	(0.0)	-	-
13	Lambayeque	2	0	(0.0)	-	-	5	0	(0.0)	-	-
14	Ica	2	0	(0.0)	-	-	1	0	(0.0)	-	-
15	Tumbes	0	0	(0.0)	-	-	2	0	(0.0)	-	-
	Total	199	15	(7.5)	8	(4.0)	118	14	(11.9)	1	(0.8)

Table 1. Distribution of colistin resistance and mcr-1 gene detection among Escherichia coli and Klebsiella pneumoniae isolates in 15 hospitals in 12 regions of Peru.

most cases, it is reserved for those bacteria that are resistant to carbapenems. Yauri *et al.* showed that 15.2% of extended-spectrum beta-lactamase-producing enterobacteria were positive for detection of the *mcr*-1 gene in a hospital in Lima ⁽⁶⁾. In our study, all isolates carrying the *mcr*-1 gene were susceptible to carbapenems.

In conclusion, the presence of *Enterobacteriaceae* species carrying the *mcr*-1 gene causing bloodstream infections distributed in several hospitals and regions of Peru, alerts about the dissemination of resistance to this last-resort antibiotic in our country. Other chromosome-mediated mechanisms of colistin resistance should be explored since we found that the *mcr*-1 gene was present in less than one third of colistin-resistant bacteria.

REFERENCES

- Mayta-Barrios M, Ramirez-Illescas J, Pampa-Espinoza L, Yagui-Moscoso M. Caracterización molecular de carbapenemasas en el Perú durante el 2019. Rev Per Med Exp Salud Publica. 2021;38(1):113–8. doi: 10.17843/rpmesp.2021.381.5882.
- Clinical and Laboratory Standards Institute. (2018). Performance standards for antimicrobial susceptibility testing; 33rd ed. CLSI standard M100. Wayne, PA: CLSI; 2021.
- El-Sayed Ahmed MAEG, Zhong LL, Shen C, Yang Y, Doi Y, Tian, GB. Colistin and its role in the era of antibiotic resistance: an extended review (2000–2019). Emerg Microbes Infect. 2020;9(1):868–85. doi: 10.1080/22221751.2020.1754133.
- 4. Krapp F, García C, Hinostroza N, Astocondor L, Rondon CR, Ingelbeen

Acknowledgments. To all the collaborators of the 15 hospitals that participated in the study.

Author contributions. All authors declare that they meet the authorship criteria recommended by the ICMJE.

Roles according to CRediT. CG and FK contributed to the conception, study design, writing and editing of the manuscript. LA and NH performed the laboratory analysis and contributed to the writing of the article. JJ contributed to the conception of the study.

Conflicts of interest. The authors declare that there is no conflict of interest.

Funding. This research was funded by the Directorate of Development Cooperation (DGD) of Belgium through a collaboration between the Antwerp Institute of Tropical Medicine and the Alexander von Humboldt Institute of Tropical Medicine.

B, et al. Prevalence of antimicrobial resistance in Gram-Negative bacteria bloodstream infections in Peru and associated outcomes: VIRAPERU Study. Am J Trop Med Hyg. 2023; Sep 18:tpmd220556. doi: 10.4269/ ajtmh.22-0556.

- Liu YY, Wang Y, Walsh TR, Yi LX, Zhang R, Spencer J, et al. Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. Lancet Infect Dis. 2016;6(2):161–8. doi: 10.1016/S1473-3099(15)00424-7.
- Yauri-Condor K, Apestegui MZ, Sevilla-Andrade CR, Sara JP, Espinoza CV, Taboada WV, et al. Extended-spectrum beta-lactamase producing enterobacterales carrying the mcr-1 gene in Lima, Peru. Rev Per Med Exp Salud Publica. 2020;37(4):711–5. doi: 10.17843/rpmesp.2020.374.5832.